Electronic Supplementary Material

Biomineralization-inspired copper-cystine nanoleaves capable of laccase-like catalysis for the colorimetric detection of epinephrine

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Fig. S1 The EDX spectrum of Cu-Cys NLs



Fig. S2 The Cu LMM Auger spectrum (a) and S 2p XPS spectrum (b) of Cu-Cys NLs



Fig. S3 The linear Lineweaver-Burk plots for Cu-Cys NLs

Catalyst	K_m /mM	<i>V_{max}</i> /(μM min ⁻¹)	k_{cat}/K_m /(×10 ⁻³ g ⁻¹ min ⁻¹ L)	Reference
Cu-Cys NLs	0.14	1.44	102.86	This work
Laccase	0.08	1.97	121.25	This work
Laccase	0.40	3.51	54.84	Wang <i>et al</i> . [1]
Laccase	0.41	6.41	156.34	Wang <i>et al.</i> [2]
Laccase	0.062	5.81	937.10	Shams et al. [3]
Laccase	0.65	0.15×10 ³	2307.69	Liang <i>et al</i> . [4]

Table S1 Kinetic parameters for Cu-Cys NLs and some reported natural laccases



Fig. S4 XRD analysis of Cu-Cys NLs (a) and ICP analysis of supernatant (b) after 6 days of incubation under different conditions



Fig. S5 The activity contribution of Cu-Cys NLs in sediment and the free Cu(II) ions in supernatant

Table S2 Comparison of laccase mimics and laccase for the quantitative determination of

epinephrine						
Method	Catalyst	Linear range /µM	LOD /µM	Reference		
Colorimetry	CH-Cu	22.8-227.6	1.4	Wang <i>et al</i> . [2]		
Colorimetry	CuCl2	20-500	10	Sivasankaran <i>et al</i> . [5]		
Amperometry	MXene/GCPE	0.02-10, 10-100	0.009	Shankar <i>et al</i> . [6]		
Amperometry	Laccase-OE	3.0-100	3.0	Molinnus et al. [7]		
Fluorimetry	GQDs	1–200	0.5	Wang <i>et al</i> . [8]		
Colorimetry	Cu-Cys NLs	9-455	2.7	This work		