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2 **Materials and Methods**

3 **Aorta intima RNA isolation**

4 Aorta intimal RNA was obtained as previously described [2]. Briefly, mouse aortas
5 were exposed, and the surrounding tissues were removed carefully. After perfusion
6 with saline, the aorta was collected and transferred to a dish containing ice-cold PBS.
7 To obtain samples from predilection sites and nonpredilection sites, the aorta was cut
8 into two parts: the aortic arch (predilection sites) and the thoracoabdominal aorta
9 (nonpredilection sites). The preparation was quickly flushed with TRIzol reagent
10 (Invitrogen, Carlsbad, CA, USA) using an insulin syringe, and the eluate was
11 collected in a 1.5-ml tube and prepared for RNA extraction. The aorta leftover (media
12 +adventitia) was stored at -80°C until RNA extraction.

13 **RNA Sequencing**

14 Total RNA from pre-treated HAECs was extracted by using Trizol reagent, and
15 purified with RNAClean XP Kit (Cat A63987, Beckman Coulter, Inc. Kraemer
16 Boulevard Brea, CA, USA) and RNase-Free DNase Set (Cat#79254, QIAGEN,
17 GmbH, Germany). The purity and integrity of total RNA samples were determined
18 using the Agilent Bioanalyzer (Agilent Technologies, Santa Clara, CA, USA). Next,
19 samples were treated with RiboZero rRNA Removal Kit (Epicentre, WI, USA) for
20 deleting rRNA. Then, rRNA-depleted and RNase R-digested RNA samples were
21 fragmented, and reverse transcribed to cDNAs with random primers. After
22 purification, cDNA libraries were quality controlled and sequenced by HiSeq2000
23 (Illumina, San Diego, CA, USA).

24 **Sequence Data Analysis**

25 Raw reads in fastq format were screened, and the adaptor sequences and low-quality
26 reads were deleted. Stringent filtering criteria were selected to diminish the ratio of
27 sequencing errors. Briefly, bases with a 3 terminal quality score lower than 10 and a
28 read-length shorter than 20 bp were removed. Genome mapping was performed based
29 on the *E. necatrix* Houghton strain reference genome sequence (GCA_000499385.1)
30 using Tophat (version 2.0.9) with a spliced-mapping algorithm. After genome
31 mapping, reads with less than two-base mismatches and multi hits ≤ 2 were retained.

32

33 **Result**

34 **lncR-GAS5 transfection efficacy and atherosclerotic model**

35 To investigate the role of lncR-GAS5 in atherogenesis, 8-week-old athero-prone
36 *ApoE*^{-/-} mice fed with a high-fat diet (HFD) were intravenously injected with
37 recombinant lentivirus vector-expressing lncR-GAS5 (Lv-GAS5), empty vector
38 (Lv-null) as a negative control, equal volume of saline fed with a HFD or a chow diet
39 (CD) as normal for 12 weeks (Supplementary Fig. 1A). As shown in supplementary
40 Fig. 1B, mRNA expression of the endothelial cells (ECs) marker CD31 was enriched
41 in the intima compared with media plus adventitia. Conversely, the smooth muscle
42 cell marker smMHC was rarely detectable in the intima, manifesting high purity ECs
43 were obtained from aortic intima (Supplementary Fig. 1C). Notably, compared with
44 the intima, the lncR-GAS5 expression was lower in the media plus adventitia,
45 indicating that lncR-GAS5 is more abundant in ECs (Supplementary Fig. 1D). We
46 also observed that lncR-GAS5 was dominantly increased in ECs of *ApoE*^{-/-} mice fed
47 with HFD (Supplementary Fig. 1E). Our result indicated that lncR-GAS5
48 overexpression repressed the mRNA level of MAP1LC3B and increased the P62
49 expression in endothelium of en face aortas (Supplementary Fig. 1F and 1G).

50 **lncR-GAS5 potentiates mice to develop abnormal hyperlipidemia, but not Alters** 51 **body weight (BW), Blood pressure, Glucose metabolism**

52 We next collected blood samples by retro-orbital venous plexus puncture to assess the
53 alteration of circulating metabolic parameters after mice were fasted for 12–14 h.
54 Result indicated that lncR-GAS5 overexpression markedly elevated expression of
55 total cholesterol (TC), triglycerides (TG), low-density lipoprotein cholesterol
56 (LDL-C), and reduced the level of high-density lipoprotein cholesterol (HDL-C)
57 under fasting conditions, respectively (Supplementary Fig. 2A, 2B, 2C and 2D). Next,
58 Body weight (BW) was further measured weekly during 12 weeks in mice fed with
59 HFD. Result indicated that even Lv-GAS5 treatment could increase BW in a time
60 dependent manner, while showed no statistically significant (Supplementary Fig. 2E).
61 Abnormal of blood pressure and blood glucose are one of the leading causes of
62 atherosclerosis. It is documented that high level of lncR-GAS5 had no influence on
63 systolic blood pressure (SBP, mmHg) in mice fed chow diet (CD) or HFD

64 (Supplementary Fig. 2F). Insulin sensitivity decrease or insulin resistant is one of the
65 main reasons of atherogenesis [1, 3]. Our result showed that Lv-GAS5 upregulation
66 had little influenced on insulin sensitivity and glucose homeostasis during an insulin
67 tolerance test (ITT) and glucose tolerance tests (GTT), respectively (Supplementary
68 Fig. 2G and 2H).

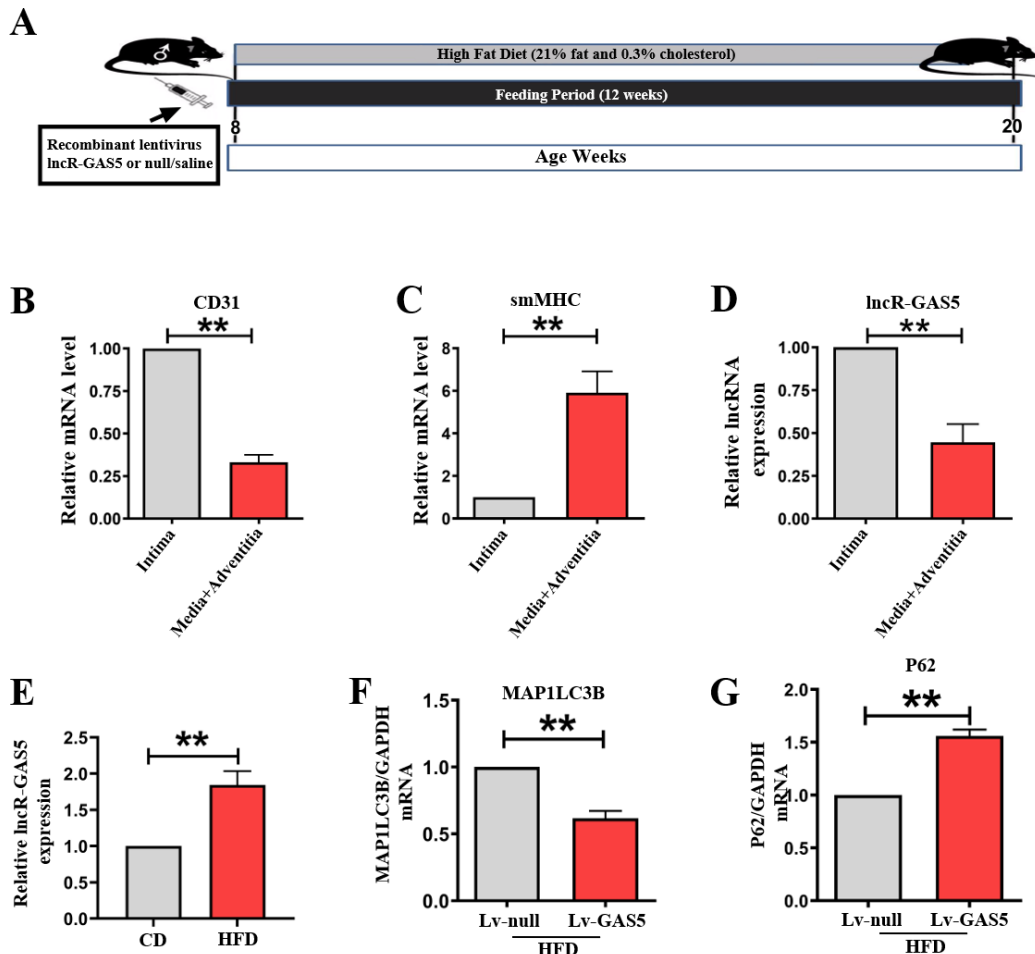
69 **lncR-GAS5 associated regulator was screened**

70 To gain insight into the molecular mechanism by which lncR-GAS5 regulates
71 autophagy, we performed tailored RNA sequencing (RNA-seq). After cells were
72 transfected with small interfering RNA against lncR-GAS5 for 24 h, the cells were
73 collected for analysis using RNA-seq. Gene ontology analysis indicated that
74 lncR-GAS5 knockdown resulted in the upregulation of 305 genes and downregulation
75 of 138 genes based on the criterion of a ≥ 2 -fold change in expression, and these
76 differentially expressed genes have been implicated in biological regulation, cellular
77 processes, , metabolic processes, and molecular function regulation (Supplementary
78 Fig. 3A). Volcano plots show the distribution of these differentially expressed genes
79 (Supplementary Fig. 3B). KEGG pathway enrichment analyses showed that the
80 differentially expressed genes are associated with immune system, lipid metabolism
81 and signal transduction (Supplementary Fig. 3C).

82 **Figure legend**

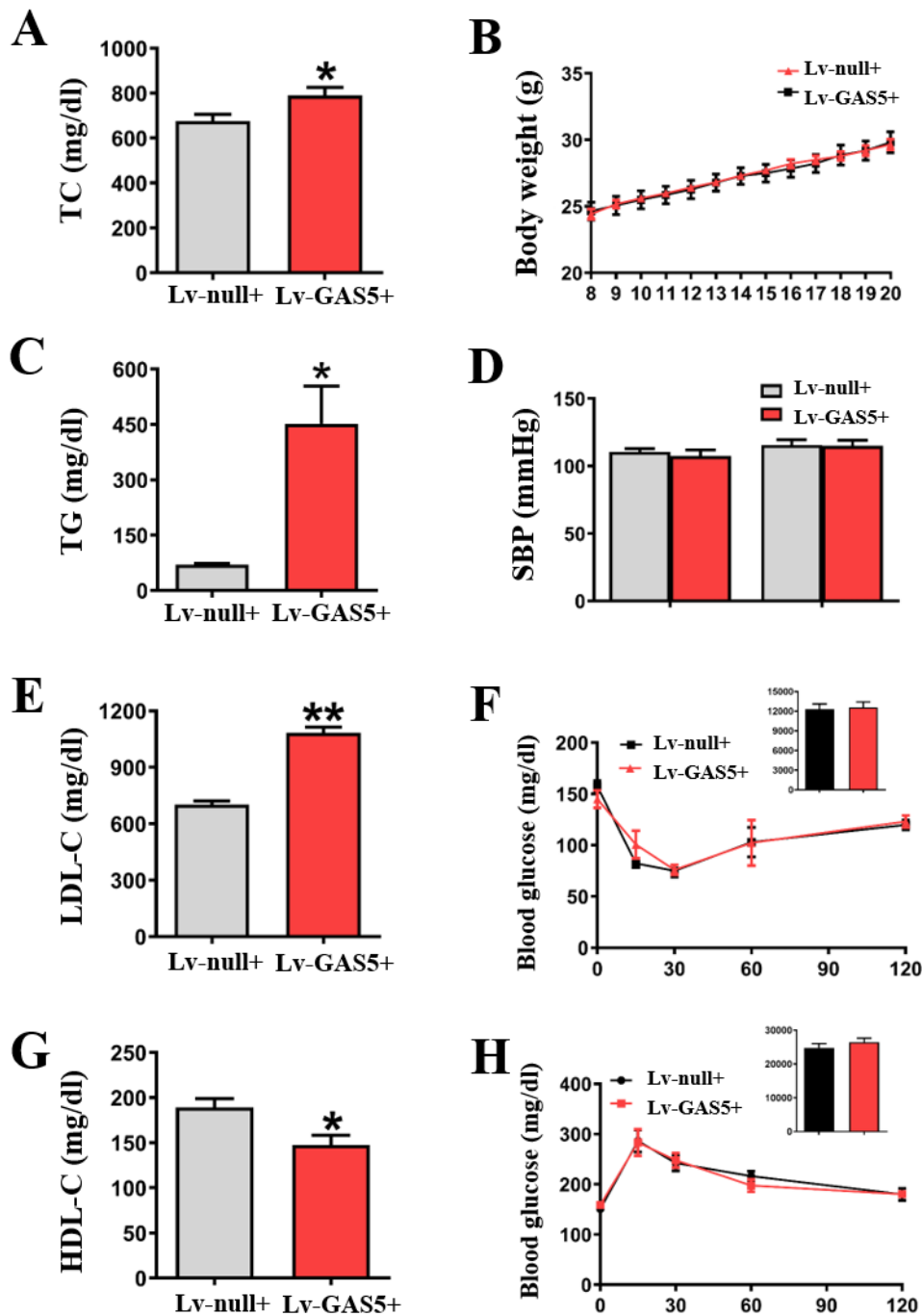
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84 **Fig. S1. Detection of lncR-GAS5 lentivirus infective efficacy and its effects on**
85 **MAP1LC3B and P62 expression in HFD-fed *ApoE*^{-/-} mice.** (A) Experiment
86 procedure. (B-D) The expression levels of CD31 (B), smMHC (C), and lncR-GAS5 in
87 HFD-fed *ApoE*^{-/-} mice. n=4 male *ApoE*^{-/-} mice. (E) The expression levels of
88 lncR-GAS5 in *ApoE*^{-/-} with HFD or CD. n=5 male *ApoE*^{-/-} mice. (F and G) The effect
89 of lncR-GAS5 on the expression of MAP1LC3B (F) and P62 (G) in HFD-fed *ApoE*^{-/-}
90 mice. ***P* <0.01 vs. Intima or CD or Lv-null group. All values are expressed as the
91 mean \pm SEM.



92

93 **Fig. S2. Fig. S2. The effect of IncR-GAS5 overexpression on lipid and lipid**
 94 **protein expression, systolic blood pressure (SBP), and blood glucose in HFD-fed**
 95 ***ApoE*^{-/-} mice.** (A-D) The expression levels of TC (A), TG (B), LDL-C (C), and
 96 HDL-C (D). (E) The body weight of mice. (F) The effect of IncR-GAS5
 97 overexpression on SBP of HFD-fed *ApoE*^{-/-} mice. (G and H) The effect of IncR-GAS5
 98 overexpression on ITT (G) and GTT (H) of HFD-fed *ApoE*^{-/-} mice. “+” indicated mice
 99 fed with HFD. n=8 male *ApoE*^{-/-} mice. **p* < 0.05, ***p* < 0.01 vs. Lv-null+ group. All
 100 data represent the mean ±S.E.M.



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103 **Fig. S3. GO and KEGG enrichment analysis of differentially expressed**
 104 **genes after lncR-GAS5 knockdown.** (A) GO enrichment analysis of the
 105 differentially expressed genes after lncR-GAS5 knockdown. (B) The distribution of
 106 differentially expressed genes after lncR-GAS5 knockdown. (C) KEGG enrichment
 107 analysis differentially expressed genes after lncR-GAS5 knockdown. n=3 batches of
 108 cells.

